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Antibody indices of infectious pathogens from serum and cerebrospinal fluid in patients with schizophrenia spectrum disorders

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Abstract

Introduction: Infectious and immunological theories of schizophrenia have been discussed for over a century. Contradictory results for infectious agents in association with schizophrenia spectrum disorders (SSDs) were reported. The rationale of this study was to investigate intrathecal antibody synthesis of the most frequently discussed neurotropic pathogens using a pathogen-specific antibody index (AI) in patients with SSD in comparison to controls.

Methods: In 100 patients with SSD and 39 mentally healthy controls with idiopathic intracranial hypertension (IIH), antibodies against the herpesviruses EBV, CMV, and HSV 1/2 as well as the protozoan *Toxoplasma gondii*, were measured in paired cerebrospinal fluid (CSF) and serum samples with ELISA-kits. From these antibody concentrations the pathogen-specific AIs were determined with the assumption of intrathecal antibody synthesis at values > 1.5.

Results: No significant difference was detected in the number of SSD patients with elevated pathogen-specific AI compared to the control group. In a subgroup analysis, a significantly higher EBV AI was observed in the group of patients with chronic SSD compared to patients with first-time SSD diagnosis ($p = 0.003$). In addition, two identified outlier EBV patients showed evidence for polyspecific immune reactions (with more than one increased AI).

Conclusions: Evidence for the role of intrathecal EBV antibody synthesis was found in patients with chronic SSD compared to those first diagnosed. Apart from a possible infectious factor in SSD pathophysiology, the evidence for polyspecific immune response in outlier patients may also suggest the involvement of further immunological processes in a small subgroup of SSD patients.

Keywords: EBV, CMV, HSV, Toxoplasmosis, Antibody index, Schizophrenia, Psychosis, SSD, Autoimmune

Introduction

Schizophrenia is a severely disabling mental illness [29]. Due to the heterogeneous presentation of clinical manifestations and biological parameters, one can assume various interrelated etiologies of schizophrenia

spectrum disorders (SSDs) [49]. Rarely, the etiology can be identified as in autoimmune psychosis caused by well-characterized antibodies like those against the anti-N-Methyl-d-aspartate receptor (NMDA-R) [35]. In spite of intensive research, other causes or underlying pathomechanisms remain widely elusive. In addition to a distinct genetic component [42], infectious [5] and immunological causes [15, 39] are frequently discussed. Among neurotropic infectious agents that have been associated most consistently with schizophrenia are *Toxoplasma gondii* as well as herpesviridae of the herpes

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simplex group (HSV1/2), cytomegalovirus (CMV), and the Epstein-Barr virus (EBV) [5, 63].

To test for exposure to infectious agents in patients with mental illnesses, various methods measuring antigens, antibodies, and nucleic acids (DNA/RNA) have been applied with mixed results [4]. Regarding antibody measurements, pathogen-specific antibodies were often detected in serum, rarely in cerebrospinal fluid (CSF), and seldom in both. Of the latter, titers have been mainly measured, of which a CSF/serum quotient was calculated. However, analyses of CSF/serum titer quotients lack sensitivity in detection of intrathecal antibody synthesis [38, 47]. This, in contrast, can be achieved with calculation of the specific antibody index (AI) in paired CSF/serum pathogen specific antibody concentrations, which allows for the identification of pathologically elevated brain-derived antibody fractions [38]. The potential elevation of AIs without further traces of infections, is a promising investigative marker for the association of infections with SSDs [38, 47]. Furthermore, several elevated AIs as polyspecific immune reactions can indicate immunological processes in the central nervous system (CNS; this has been reported in multiple sclerosis for MRZ reaction) [20].

Nonetheless, no study could be identified investigating intrathecal antibody synthesis against *T. gondii* and the Herpesviridae HSV1/2, CMV, and EBV using pathogen-specific AIs in SSDs. Therefore, the rationale of this study was to investigate the fractions of brain-derived antibody synthesis of specific antibodies using AIs in patients with SSD and compare them with a mentally healthy control group to test the association of the four aforementioned infectious agents with SSDs. Furthermore, several elevated AIs should provide insight into polyspecific immune reactions as possible inflammatory signs of the CNS, investigating immunological causes of schizophrenia.

Methods

Ethical approval for the current study was provided by the local ethics committee in the context of a larger retrospective research project (Medical Faculty of the University of Freiburg, EK-Fr 609/14). Patients with SSDs gave their written informed consent before lumbar puncture. The control patients were contacted retrospectively and asked for their consent to use the residual serum/CSF material for research purposes.

Study sample

In the patient group, 100 consecutive patients with SSD treated as inpatients in the Department of Psychiatry and Psychotherapy of the Medical Center of the University of Freiburg were included. Diagnoses and comorbidities

were established by experienced senior consultant psychiatrists according to ICD-10 criteria and extracted for this study through chart review. Exclusion criteria were defined as known substance use disorders and immunological disorders known for brain involvement. All patients were offered a magnetic resonance imaging (MRI) scan of the brain and an electroencephalography (EEG) as part of the routine workup. During the admission routine, comprehensive demographic and clinical data are collected. In addition, psychometric scales, such as the Global Assessment of Functioning (GAF) [2], Clinical Global Impression (CGI) [41], and psychopathological scores based on guidelines published by the German Association for Methodology and Documentation in Psychiatry (AMDP) [9] were assessed.

A mentally healthy control group consisted of 39 patients with idiopathic intracranial hypertension (IIH). The exclusion criteria for the control group were as follows: secondary forms of intracranial hypertension and psychiatric or other neurological disorders (except headaches). The control group was already published in earlier articles [27, 31, 39, 40].

Antibody index

AI enables the detection of the intrathecal synthesis of a pathogen-specific antibody by distinguishing between antibodies originating in the brain and antibodies that passively cross the blood-CSF barrier (BCSFB) [38, 47]. This is mathematically possible by relating the specific CSF/serum antibody quotient (Q_{spec}) to the CSF/serum quotient of total IgG antibodies (Q_{IgG} ; hence, $\text{AI} = Q_{\text{spec}}/Q_{\text{IgG}}$) [38]. However, because polyspecific increased intrathecal IgG production would lead to an underestimation of pathogen-specific antibodies, AI is calculated in reference to Q_{lim} if the total IgG exceeds Q_{lim} : $\text{AI} = Q_{\text{spec}}/Q_{\text{lim}}$ (if $Q_{\text{IgG}} > Q_{\text{lim}}$) [38]. All IgG antibody measurements for EBV capsid antigen, HSV1/2, CMV, and *T. gondii* in serum and CSF were performed using commercially available enzyme-linked immunosorbent assay (ELISA) kits, according to the manufacturer's instructions (Euroimmun®, Lübeck, Germany). Total IgG was determined nephelometrically using an Atellica NEPH 630 (Siemens Healthineers®, Erlangen, Germany) as specified by the manufacturer. Paired serum and CSF samples from the same day were measured consecutively on the same ELISA plate, side by side, to avoid inter-assay variation. ELISA standard curves for all parameters ranged from 5–100 Units (U; 230 in the case of HSV), so that a cut-off value of < 5 U was defined, at which the measurements were not considered positive. AI was calculated only if measurable antibody concentrations were found in CSF and serum. An AI between 0.7–1.5 was considered normal, and all values above this

were considered positive for intrathecal antibody synthesis. All AIs below 0.7 were measured again and checked for plausibility. Since the ELISA kits were optimized for calculating AIs, group comparisons of the serum/CSF concentrations are possible, but no direct statement on total IgG values.

CSF routine parameters and anti-neuronal antibodies

Routine CSF analyses and testing for anti-neuronal antibodies were conducted in the CSF laboratory of the Clinic of Neurology and Neurophysiology at the University Hospital Freiburg, which was described in earlier publications [15, 20]. For CSF antibodies against neuronal cell surface antigens (NMDA-R, LGI1, etc.), fixed cell-based assays were performed by Euroimmun® (Lübeck, Germany). For serum autoantibodies against paraneoplastic intracellular antigens (Yo, Hu, etc.) immunoblots by Ravo Diagnostika® (Freiburg, Germany) were performed [15]. The remaining CSF was preserved at -80°C after routine analysis for further analysis.

Statistical analyses

Statistical analyses of the acquired data were performed using the SPSS Version 27 (IBM, Armonk, USA). Group differences of patients and controls as well as subanalyses of patient subgroups for categorical variables (e.g., number of increased AIs) were statistically assessed by Chi-squared tests. For continuous variables (e.g., antibody concentrations), normality, assessed by the Shapiro–Wilk test, could only be assumed for a few variables. Therefore, a nonparametric Mann–Whitney U test for independent samples was conducted for all continuous variables. Secondary analysis with correction of sex was performed for continuous variables with an analysis of covariance (ANCOVA) and for categorical variables with binary logistic regression. Spearman's rank correlation coefficient was used for the correlation between AIs and CSF

routine parameters, age, psychometric scales, the number of suicide attempts and previous inpatient stays. Due to the descriptive nature of our study and the explorative nature of the subanalyses no correction for multiple testing was conducted. In all statistical analyses, a p-value lower than 0.05 was presumed statistically significant. All patients with antibody indices higher than two standard deviations above the mean value of the corresponding study group were characterized in more detail. Data visualization was realized with R package ggplot2 [37, 57] and Adobe Illustrator (Adobe Inc., San José, CA).

Results

Demographic data

In 100 patients with SSD and 39 controls, no significant difference in age ($z = -0.516$, $p = 0.606$) could be identified. Predominantly, women were included in the patient group (60% women vs. 40% men) as well as the control group (85% women vs. 15% men), with a significant sex difference between the two groups ($\chi^2 = 7.678$, $p = 0.006$). When examining the patient group with SSD, paranoid schizophrenia (56%) with chronic or recurrent courses (58%) was mainly diagnosed. At the time of lumbar puncture, only three patients (3%) did not receive any psychiatric medication. All clinical and demographic data are presented in Additional file 2: Table S1.

Pathogen specific antibody index

Several patients with elevated pathogen-specific AIs, indicating intrathecal IgG synthesis, were identified in the patient and control groups (Table 1 and Fig. 1). When comparing the number of patients with elevated AIs between SSD patients and controls, no significant differences were observed for any of the following pathogens: EBV, CMV, HSV, and *T. gondii*. For *T. gondii*, there is even a trend of frequently increased AIs in the control group ($p = 0.061$), and it is the only pathogen for which

Table 1 Number of participants with elevated pathogen specific antibody indices (AIs)

	Elevated AIs in SSD patients (number of measurable AIs)	Elevated AIs in IHH controls (number of measurable AIs)	Statistics
EBV	4 (n = 84)	1 (n = 37)	$\chi^2 = 0.275$, $p = 0.600$
CMV	8 (n = 31)	1 (n = 15)	$\chi^2 = 1.343$, $p = 0.246$
HSV	5 (n = 59)	5 (n = 26)	$\chi^2 = 2.011$, $p = 0.156$
<i>T. gondii</i>	0 (n = 17)	3 (n = 16)	$\chi^2 = 3.506$, $p = 0.061$
Number of elevated AIs			$\chi^2 = 2.942$, $p = 0.451$
0	85 (85%)	32 (82%)	
1	13 (13%)	5 (13%)	
2	2 (2%)	1 (3%)	
3	0 (0%)	1 (3%)	

AI antibody index, SSD schizophrenia spectrum disorder, IHH idiopathic intracranial hypertension, EBV Epstein–Barr virus, CMV human cytomegalovirus, HSV herpes simplex virus, *T. gondii* *Toxoplasma gondii*

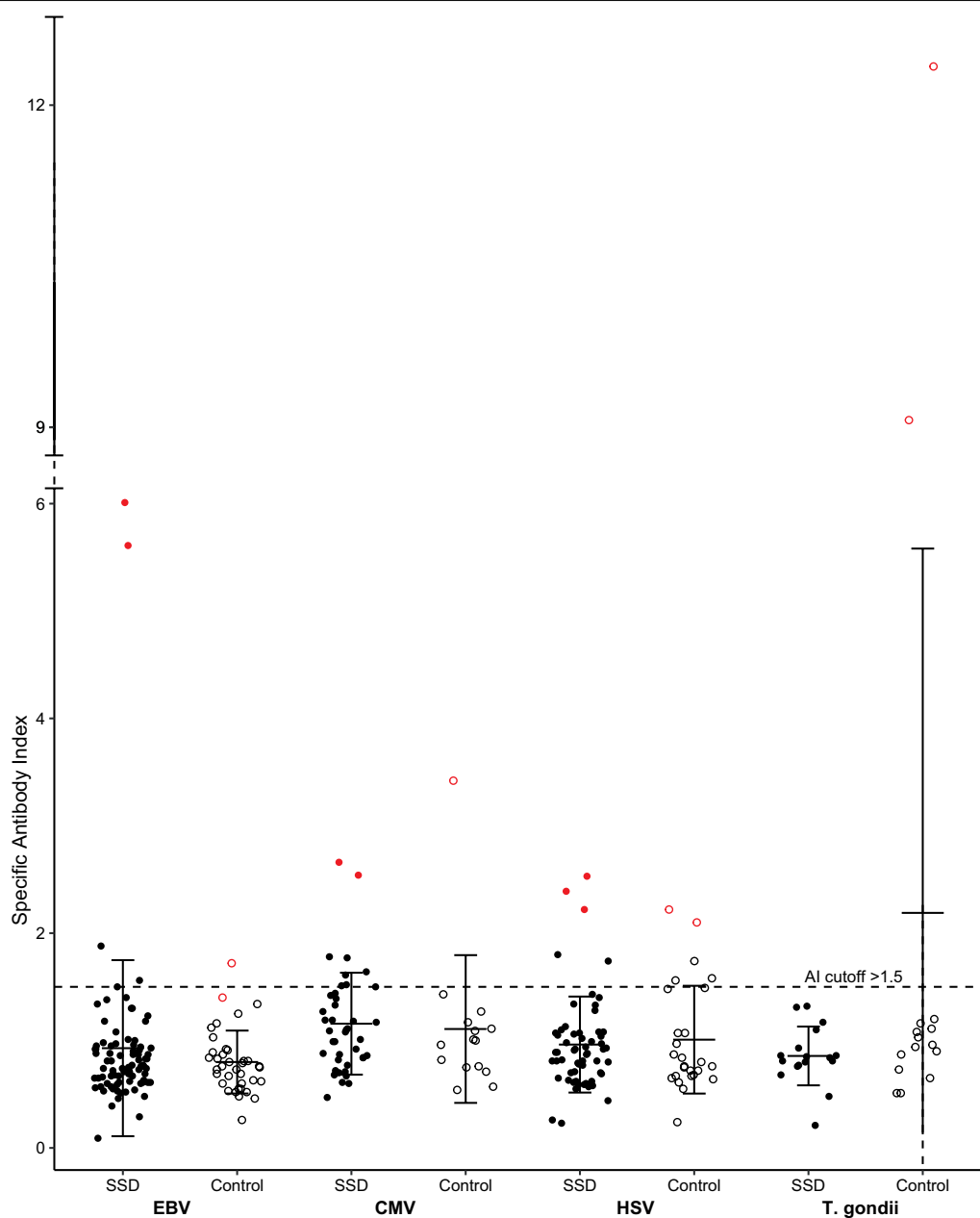
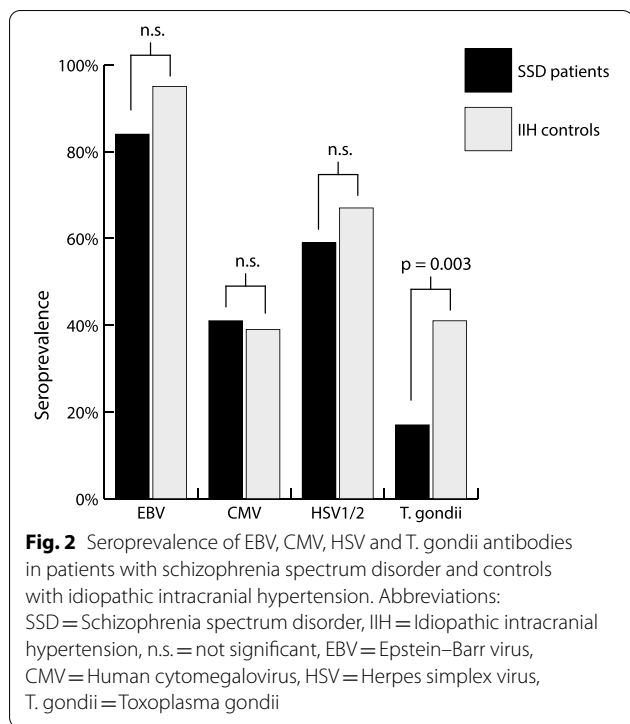


Fig. 1 Specific antibody indices of the pathogens EBV, CMV, HSV and *T. gondii* in patients with schizophrenia spectrum disorder and controls with idiopathic intracranial hypertension. Error bars indicate mean value \pm one standard deviation. All datapoints in red indicate outliers of more than 2 standard deviations over mean value (outlier patients are presented in detail in Sect. 3.6). Abbreviations: AI = Antibody Index, SSD = Schizophrenia spectrum disorder, EBV = Epstein–Barr virus, CMV = Human cytomegalovirus, HSV = Herpes simplex virus, *T. gondii* = *Toxoplasma gondii*

no elevated AI was found in the SSD group. There was also no significant difference in the number of patients with more than one elevated AI ($p = 0.451$). When applying higher thresholds for elevated AIs of more than two [10, 21], the group comparison of the number of patients with elevated AIs remained statistically insignificant. In a secondary analysis with correction for sex, the differences

remained insignificant, and the tendency for an increased number of elevated *T. gondii* AIs in IiH controls could no longer be detected (Wald < 0.001 , $p = 0.999$).

When assessing the number of patients with measurable antibodies, the EBV showed the highest seroprevalence, with up to 95% in the control group (Fig. 2). The seroprevalence of herpesviruses is similar between the



two study groups, while for *T. gondii* the fraction of seropositive SSD patients is with 17% less than half the fraction of patients in the control group with 41% ($p = 0.003$). After correction for sex, this effect remained significant ($\beta = -1.159$, Wald = 7.171, $p = 0.007$).

Upon comparison of individual antibody concentrations in CSF and serum, a statistical difference between the CSF to serum ratios for EBV ($p = 0.006$) and CMV ($p = 0.014$) was striking (Table 2). However, when this quotient of pathogen-specific antibody concentrations from CSF and serum was related to the total antibody concentration and the blood-CSF function (i.e., the AI was calculated), no significant differences were observed. The results of the significantly higher CSF to serum ratios for EBV and CMV are further described and discussed in the Additional file 1.

Routine cerebrospinal fluid diagnostics and neuronal autoantibody findings

In CSF routine diagnostics, the SSD group showed significantly higher total protein concentrations ($p < 0.001$) with significantly more patients with elevated protein levels than the control group ($p = 0.010$). AQs were also significantly elevated in patients with SSD ($p < 0.001$). After

Table 2 Results of antibody measurement and seroprevalence

Antibody levels (Mean ± SD)	Patients with schizophrenia spectrum disorders (n = 100)*	IIH Controls (n = 39)*	p-value
EBV	n = 84 (84%)	n = 37 (95%)	Chi ² = 2.942 p = 0.086
Serum (U)	51,574.59 ± 50,164.67	54,007.71 ± 47,090.30	z = -0.602 p = 0.547
CSF (U)	120.73 ± 155.75	79.02 ± 81.99	z = -0.875 p = 0.382
CSF/Serum	2.81 ± 4.19	1.57 ± 0.84	z = -2.768 p = 0.006
AI	0.93 ± 0.82	0.80 ± 0.29	z = -0.535 p = 0.593
CMV	n = 41 (41%)	n = 15 (39%)	Chi ² = 0.075 p = 0.784
Serum (U)	27,638.94 ± 27,336.05	33,900.25 ± 33,022.25	z = -0.916 p = 0.360
CSF (U)	64.14 ± 55.43	66.55 ± 77.03	z = -0.139 p = 0.890
CSF/Serum	2.64 ± 1.05	2.16 ± 1.69	z = -2.470 p = 0.014
AI	1.16 ± 0.47	1.11 ± 0.69	z = -1.018 p = 0.309
HSV1/2	n = 59 (59%)	n = 26 (67%)	Chi ² = 0.694 p = 0.405
Serum (U)	62,392.74 ± 33,027.48	70,727.88 ± 44,320.81	z = -0.019 p = 0.985
CSF (U)	160.16 ± 197.33	155.11 ± 130.48	z = -0.048 p = 0.962
CSF/Serum	2.77 ± 3.93	2.17 ± 1.52	z = -0.992 p = 0.321
AI	0.96 ± 0.45	1.01 ± 0.50	z = -0.105 p = 0.916
T. gondii	n = 17 (17%)	n = 16 (41%)	Chi ² = 8.946 p = 0.003
Serum (U)	21,313.23 ± 13,672.51	27,006.46 ± 14,736.33	z = -1.513 p = 0.136
CSF (U)	47.60 ± 42.40	64.54 ± 39.71	z = -1.693 p = 0.094
CSF/Serum	2.31 ± 1.29	4.33 ± 6.46	z = -0.036 p = 0.986
AI	0.86 ± 0.27	2.91 ± 3.40	z = -1.549 p = 0.127

Significant p-values are marked in bold. EBV Epstein-Barr virus

U unit, CSF cerebrospinal fluid, CMV human cytomegalovirus, HSV herpes simplex virus, *T. gondii* *Toxoplasma gondii*, SD standard deviation, IIH idiopathic intracranial hypertension, AI antibody index

* Only measurable antibody levels were analyzed

correction for sex in secondary analysis, the significant difference in total protein concentrations ($F_{(1,136)}=6.588$, $p=0.011$), number of patients with elevated total protein levels ($\beta=1.074$, Wald=4.625, $p=0.032$), and AQ ($F_{(1,136)}=4.970$, $p=0.027$) persisted.

One patient presented with questionable anti-NMDA-R IgG autoantibodies in serum (negative in CSF; only tested in N=74 patients). No autoantibodies against intracellular antigens could be found (N=86; Additional file 2: Tables S2, S3).

Instrumental diagnostics

Instrument-based diagnostics revealed MRI alterations in 63% of patients with SSD and EEG abnormalities in 31% of patients with SSD. Frequent abnormalities were non-specific white matter changes in MRI (28%) and intermittent generalized slowing in EEG (25%; see Additional file 2: Table S4).

Correlation analyses

In the correlation analysis, EBV AI correlated significantly with disturbances of orientation ($r=0.253$, $p=0.042$; N=65). HSV AI correlated with delusions ($r=0.363$, $p=0.015$; N=44). For age, a negative correlation with CMV AI ($r=-0.383$, $p=0.014$; N=41) could be detected. Negative correlations for Toxoplasma AI for ego-boundary disturbances ($r=-0.637$, $p=0.026$; N=12) and hallucinations ($r=-0.826$, $p=0.001$; N=12) could be found. EBV and HSV AI correlated with each other ($r=0.391$, $p=0.004$; N=53).

Characteristics of AI outliers and subanalyses

All outlier patients with AIs two standard deviations over the mean value are marked red in Fig. 1. Interestingly, when counting the borderline positive CMV AI in Patient 3 (Table 3), half of the patients with outlier AIs showed two elevated AIs, with Patient 1 presenting with two outlier AIs. In contrast, the CSF findings are all inconspicuous, except for oligoclonal bands (OCBs) in Patient 2, indicating an intrathecal oligoclonal IgG synthesis, and an unspecific identical OCB in CSF and serum in Patient 3, which is considered a normal finding, and may be due to the elevated AIs in these patients. Further diagnostic investigation with EEG and MRI identified only mild abnormalities in some patients. It is remarkable that the two outlier EBV patients both presented with intrathecal HSV1/2 antibody synthesis; in one of these patients, OCBs were also detected. These are indications of a polyspecific immune response rather than an acute EBV infection.

In a subgroup analysis of the 15 SSD patients with at least one elevated AI compared to the remaining 85 patients without elevated AI, no significant age or sex

difference was found. The CSF protein concentrations were lower, on average, in the group of patients with elevated AI than in the rest of the SSD patients (total protein 351.20 vs. 472.14 mg/dl; AQ 4.37 vs. 5.77), but did not differ significantly. No significant differences in EEG and MRI findings were observed.

In a further subgroup analysis between patients with first-time SSD diagnosis (n=42) and patients with chronic or relapsing SSD (n=58), no significant age or sex difference was found. When assessing the specific AIs, a significantly higher mean EBV AI was observed in patients with chronic SSD relative to patients with first-time diagnosis (EBV AI: 1.10 ± 1.07 in 45 patients with chronic SSD vs 0.73 ± 0.25 in 39 patients with first-time diagnosis; $z=-2.933$, $p=0.003$). Similarly, when comparing the number of patients with elevated EBV AI values, there was a clear tendency of increased AIs towards the group with chronic SSD (elevated EBV AI > 1.5: 4 out of 45 with chronic SSD vs. 0 out of 39 with first-time diagnosis; $\chi^2=3.640$, $p=0.056$). No significant differences were observed for the other pathogens or the number of patients with multiple elevated AIs.

Discussion

In this study, the difference between SSD patients and controls regarding intrathecal antibody synthesis from the specific pathogens EBV, CMV, HSV, and *T. gondii* was investigated, as an association with schizophrenia has been shown in previous studies [4, 59]. Nevertheless in this study, no significant difference was found between the two groups in the number of patients with elevated pathogen-specific AI or in the number of patients with multiple elevated AIs. Two patients with SSDs displayed polyclonal antibody synthesis with elevated AIs for EBV and HSV. Interestingly, there were higher EBV AI levels in the group of patients with chronic SSD compared to patients who were diagnosed with SSD for the first time.

EBV

Of all pathogen-specific antibodies, those against EBV were the most frequently detected in the serum of patients and controls. This is not surprising, since EBV has a known high seroprevalence in healthy populations with 73–95%, and in patients with multiple sclerosis, seroprevalence is closer to 100% [1, 6, 18]. EBV infection in childhood was associated with the occurrence of a psychotic episode in adolescence in a prospective study [23]. In several previous studies, no differences in antibody levels of SSD patients with control populations have been observed [4, 59], whereas other studies reported elevated antibodies against the EBV VCA protein and whole virion with no differences in EBV EBNA-1 antibodies [11]. Elevated levels of EBV antibodies are associated, on

Table 3 Characteristics of outlier patients with increased pathogen specific AI

Patient	Sex	Age	Psychiatric syndrome	Somatic co-morbidity	Als	Medication	CSF	Other immunological changes	MRI	EEG	Previous inpatient stays	Course of disease
Pat. (1)	F	End 40	Schizoaffective syndrome ADHD	None	EBV 6.01, HSV 2.53, CMV/T. gondii not measurable	Haloperidol, quetiapine, diazepam	Inconspicuous	None	Inconspicuous	Inconspicuous	Unknown	Chronic
Pat. (2)	M	Mid 20	Paranoid hallucinatory syndrome Depression	None	EBV 5.61, HSV 1.74, CMV/T. gondii not measurable	Olanzapine, escitalopram	Intrathecal oligoclonal IgG-synthesis (OCBs)	None	Inconspicuous	Inconspicuous	10	Chronic
Pat. (3)	M	Mid 20	Paranoid hallucinatory syndrome	None	HSV 2.39, CMV 1.5, T. gondii 0.68 EBV not measurable	Risperidone	1 identical oligoclonal band (OCB) in CSF and serum (non-specific)	None	Small developmental venous anomaly, otherwise inconspicuous	Inconspicuous	1	First diagnosis
Pat. (4)	F	End 40	Acute polymorphic psychotic disorder	Hyperthyroidism	HSV 2.22, EBV 0.98, CMV/T. gondii not measurable	Olanzapine	Inconspicuous	None	Inconspicuous	Inconspicuous	Unknown	Recurrent
Pat. (5)	F	Mid 20	Paranoid hallucinatory syndrome	Polycystic ovary syndrome, hyperandrogenemia	CMV 2.66, HSV 1.07, EBV 0.60, T. gondii not measurable	Clozapine, aripiprazole	Inconspicuous	None	Enlarged Virchow-Robin spaces	Inconspicuous	Unknown	Chronic
Pat. (6)	F	~ 20	Paranoid hallucinatory syndrome	None	CMV 2.54, EBV 1.38, HSV/T. gondii not measurable	Risperidone, promethazine	Inconspicuous	None	Pinealis cyst, small white matter lesion	Intermittent general slowing	None	First diagnosis

All elevated intrathecal pathogen-specific Als are marked in bold

F female, M male, BBB blood-brain-barrier, OCB oligoclonal bands, CSF cerebrospinal fluid, EBV Epstein-Barr virus, HSV herpes simplex virus, AI antibody index

the one hand, in combination with genetic susceptibility to schizophrenia with a greater risk for schizophrenia diagnosis [11], and on the other hand, with a lower level of cognitive performance [12]. The latter is of great interest, since in this study a correlation between the EBV-specific AI and the AMDP score for disturbances of orientation was observed. This may be a hint at greater cognitive impairment in SSD patients with an increased intrathecal synthesis of EBV antibodies. Nevertheless, controlled studies that account for the variety of confounding factors involving cognition are needed to reach a valid conclusion in this regard.

Furthermore, subanalysis revealed that elevated EBV AIs were associated with patients with chronic SSD compared to patients with first-time diagnoses independent of age and sex. There is little information available on EBV infections and the chronicity of SSDs. Although it is well-known that EBV causes latent infections in a wide variety of body regions, including the brain, with the potential to reactivate [12]. Such latent EBV infections and reactivations in the CNS have been associated with encephalitis in stem cell recipients [30] and inflammatory immune reactions in active multiple sclerosis lesions [6, 56]. With the potential for latent infections and recurrent reactivations in mind, it is imaginable that EBV may play a potential modulating role in a subgroup of patients with chronic SSD.

CMV

The association of CMV and schizophrenia is much debated, and several contradictory results emerge [52]. Increased CMV antibody concentrations in serum [32, 45, 48, 50] and CSF [28] have been described in the literature in patients with schizophrenia; however, no significant differences have been reported in other studies [19, 25, 52, 53, 62] as in this study. The AI of CMV in this study was the only studied pathogen to correlate negatively with age. Thus, younger, possibly more immunocompetent patients, appear to be more prone to intrathecal CMV antibody synthesis. Albrecht et al. (1980) found no age difference regarding CMV antibody concentrations in serum, but observed an increase in the CSF/serum ratio of CMV antibodies along with increased age-dependent blood–brain-barrier permeability. Other antibody indices, such as from the John Cunningham virus in multiple sclerosis, tend to have a rather positive correlation while increasing with age [26].

HSV

As with the other pathogens, the study evidence on HSV 1 and HSV 2 infections with schizophrenia is quite ambiguous [24, 28, 32, 33, 48, 53, 59]. Even clinically controlled trials of anti-infective therapy with valacyclovir

were conducted, but failed to show symptom improvement [8]. However, there is strong evidence that serologic detection of HSV-1 affects cognitive functioning in patients with SSD [13, 55]. In particular, memory and attentional abilities seem to be impaired [55]. No evidence for this could be found in our study, but a correlation between HSV AI and delusions was observed. This is an interesting finding, as previous studies have described an association of HSV seropositivity with negative symptoms [7].

Toxoplasmosis

A variety of studies have found an association between *T. gondii* infection and schizophrenia [28, 46, 51, 60], while others did not [24, 59, 60]. Among the latter one study reported that in areas with a low seroprevalence of *T. gondii* infections, patients with an established diagnosis of schizophrenia often have negative *T. gondii* findings, while patients with recent onset of psychosis might be associated with it [17, 60, 61]. Hence, the timing of toxoplasmosis infection with symptom onset of schizophreniform symptoms appears to play an important role [60]. In this study the proportion of patients with chronic SSD and first-time diagnosis is balanced; no difference between *T. gondii* AI could be found between these subgroups. Nevertheless, a very low seroprevalence of 17% was observed in SSD patients, which is in line with previously published negative studies. However, it is striking that the seroprevalence of the control group, with 43%, corresponds to the previously described seroprevalence of the general population in southern Germany with 44% [58]. A possible explanation may be the neuroleptic medication of the patient group, as inhibition of replication of *T. gondii* by neuroleptics has been shown in vitro [22]. Nevertheless, a previous study comparing *T. gondii* AIs from bipolar patients to patients with IHD reported no significant difference in the number of elevated AIs or seroprevalence [47]. Of the patients with SSD and available AIs, negative correlations were found for AMDP scores for ego (boundary) disturbances and hallucinations. In contrast, previous studies for *T. gondii* described seropositive patients with a higher intensity of psychotic symptoms [3].

Polyspecific immune response

Two of the patients with chronic SSD and elevated EBV AIs also had intrathecal HSV1/2 antibody synthesis, with OCBs also detected in one of them, suggesting a polyspecific immune response. This is reminiscent of the MRZ reaction, which is positive in patients with multiple sclerosis and for which at least two elevated AIs of measles, rubella, and/or varicella zoster virus AIs have to be increased [21]. Among the cases with multiple elevated

AIs detected here, patients with autoimmune psychosis could be hidden [35]. This phenomenon seems to be rare (at 2% in our cohort, 3% in the subgroup of chronic SSD). In line with this, an earlier study described a positive MRZ reaction in two of the 39 (5%) patients with SSDs [14]. Nevertheless, a similar rate of multiple positive AIs was found in our control group of IIH patients. The trigger of autoimmunity in these cases will have to be further investigated in the future.

Limitations

Individual analysis of antibody concentrations in CSF and serum revealed significant higher CSF to serum ratios for EBV and CMV (Table 2). Nevertheless, when the pathogen-specific AIs were calculated with these quotients and the CSF/serum quotient of total IgG antibodies, no significant differences were observed between SSD and control group. This finding shows the importance of calculating AIs, instead of CSF/serum antibody quotients alone, to avoid wrong conclusions. A more detailed discussion of these methodological aspects can be found in the Additional file 1.

The major limiting factor is the neurological control group of patients with IIH, as the disorder may be associated with *T. gondii* [16, 36] or was also reported to be imitated by atypical HSV-2 infection [44]. Therefore, it could underestimate the study results. Furthermore, although only control patients with no known mental illness were included, no systematic screening for mental illness or subclinical psychiatric symptoms, as prescribed in IIH [34], was performed in the control group. For ethical reasons the establishment of a healthy control group with lumbar punctures was not possible.

Furthermore, a significant sex discrepancy may have influenced the results since, for example, an influence of sex on the seroprevalence of *T. gondii* is known [58]. Therefore, secondary analyses with correction for sex were conducted. Another influential factor may be the medication of the patient group. While antibody concentrations of CMV, for instance, were unchanged under medication in previous studies [54], other studies showed lower concentrations in treated patients compared to untreated patients [28]. In the case of *T. gondii*, there is evidence that neuroleptic medication *in vitro* may directly inhibit the replication of the protozoan and thus affect antibody concentrations [22]. In addition, potential influencing factors such as rural living or pet ownership (especially cats due to their relationship with *T. gondii* [58]) are not known. Regarding methodology, statements about absolute antibody concentrations are only possible to a limited extent since the ELISA kits were designed for the measurement of pathogen-specific AIs. Although

ELISAs have improved significantly in recent years, some elaborate methods, such as reactivity to *T. gondii* proteins in the Western blot technique, appear even more sensitive in detecting seropositive patients [60]. Some AIs were <0.7 and thus could not be measured optimally (cf. Figure 1) as known from other studies [43, 47]. Finally, it must be mentioned that although the determination of pathogen-specific AIs entails many advantages, such as the detection of immunological responses in the CNS even years after pathogen contact or detection of polyspecific immune responses [47], it does not provide concrete information about infectious disease status or involvement.

Conclusion

In summary, in this assessment of pathogen-specific intrathecal antibody synthesis by AI, we did not find significant group difference between SSD patients and the control group of IIH patients. Subanalyses of SSD patients revealed a possible role of latent EBV infection in chronic SSDs with evidence for a polyspecific immune response of the CNS in 3% of patient with chronic vs. 0% with first diagnosed SSD. Given the still highly inconclusive study situation, further investigations on the influence of infectious pathogens, especially EBV, on schizophrenia using multimodal approaches, including CSF analyses, are necessary.

Supplementary Information

The online version contains supplementary material available at <https://doi.org/10.1186/s12987-022-00355-7>.

Additional file 1: Increased CSF/serum IgG ratios for EBV and CMV antibodies.

Additional file 2: Table S1. Clinical and demographic data. **Table S2.** Cerebrospinal fluid routine diagnostics. **Table S3.** Number of participants with abnormal cerebrospinal fluid diagnostics. **Table S4.** Number of magnetic resonance imaging (MRI) and electroencephalography (EEG) alterations in the patient group with schizophrenia spectrum disorders.

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Author contributions

KR wrote the paper. DE critically revised the first draft. KR, AB, BLF, LTV and DE organized the study and created the study design. AB and BLF performed the antibody measurements. RD performed the CSF routine analyses and supervised the AI calculations. AI calculations were performed by KR and AB. KR and AB performed the statistical analyses. SJM and AS supported the statistical analyses. AB, BP, AS, KN and RD revised the manuscript critically focusing on clinical and statistical aspects. All authors were critically involved in the theoretical discussion and composition of the manuscript. All authors read and approved the final manuscript.

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Availability of data and materials

All necessary information is displayed descriptively in the results section.

Declarations

Ethics approval and consent to participate

This retrospective analysis received approval from the local ethics committee of the University of Freiburg (EK Fr 609/14). The patients gave their written informed consent before the lumbar puncture.

Consent for publication

Not applicable.

Competing interests

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References

- Abrahamyan S, Eberspächer B, Hoshi M-M, Aly L, Luessi F, Groppa S, Klotz L, Meuth SG, Schroeder C, Grüter T, Tackenberg B, Paul F, Then-Bergh F, Kümpfel T, Weber F, Stangel M, Bayas A, Wildemann B, Heesen C, Zettl U, Warnke C, Antony G, Hessler N, Wiendl H, Bittner S, Hemmer B, Gold R, Salmen A, Ruprecht K. Complete Epstein-Barr virus seropositivity in a large cohort of patients with early multiple sclerosis. *J Neurol Neurosurg Psychiatry*. 2020;91:681–6. <https://doi.org/10.1136/jnnp-2020-322941>.
- American Psychiatric Association. Diagnostic and statistical manual of mental disorders: DSM-IV-TR. 4th ed. Arlington, VA: American Psychiatric Assoc; 2009.
- Amminger GP, McGorry PD, Berger GE, Wade D, Yung AR, Phillips LJ, Harrigan SM, Francey SM, Yolken RH. Antibodies to infectious agents in individuals at ultra-high risk for psychosis. *Biol Psychiat*. 2007;61:1215–7. <https://doi.org/10.1016/j.biopsych.2006.09.034>.
- Arias I, Sorlozano A, Villegas E, Luna JdD, McKenney K, Cervilla J, Gutierrez B, Gutierrez J. Infectious agents associated with schizophrenia: a meta-analysis. *Schizophr Res*. 2012;136:128–36. <https://doi.org/10.1016/j.schres.2011.10.026>.
- Benros ME, Mortensen PB. Role of infection, autoimmunity, atopic disorders, and the immune system in schizophrenia: evidence from epidemiological and genetic studies. *Curr Top Behav Neurosci*. 2020;44:141–59. https://doi.org/10.1007/7854_2019_93.
- Bjornevik K, Cortese M, Healy BC, Kuhle J, Mina MJ, Leng Y, Eledge SJ, Niebuhr DW, Scher AI, Munger KL, Ascherio A. Longitudinal analysis reveals high prevalence of Epstein-Barr virus associated with multiple sclerosis. *Science*. 2022;375:296–301. <https://doi.org/10.1126/science.abbj8222>.
- Bolu A, Oznur T, Tok D, Balıkcı A, Sener K, Celik C, Gulsun M. Seropositivity of neurotropic infectious agents in first-episode schizophrenia patients and the relationship with positive and negative symptoms. *Psychiatr Danub*. 2016;28:132–8.
- Breier A, Buchanan RW, D'Souza D, Nuechterlein K, Marder S, Dunn W, Preskorn S, Macaluso M, Wurfel B, Maguire G, Kakar R, Highum D, Hoffmeyer D, Coskinas E, Litman R, Vohs JL, Radnovich A, Francis MM, Metzler E, Visco A, Mehdiyou N, Yang Z, Zhang Y, Yolken RH, Dickerson FB. Herpes simplex virus 1 infection and valacyclovir treatment in schizophrenia: results from the VISTA study. *Schizophr Res*. 2019;206:291–9. <https://doi.org/10.1016/j.schres.2018.11.002>.
2018. Das AMDP-System: Manual zur dokumentation psychiatrischer Befunde, 10th ed., korrigierte Auflage. Hogrefe, Göttingen. ISBN: 9783844428858.
- Dersch R, Tebartz van Elst L, Hochstuhl B, Fiebich BL, Stich O, Robinson T, Matysik M, Michel M, Runge K, Nickel K, Domschke K, Endres D. Anti-thyroid peroxidase and anti-thyroglobulin autoantibodies in the cerebrospinal fluid of patients with unipolar depression. *J Clin Med*. 2020;9:2391. <https://doi.org/10.3390/jcm9082391>.
- Dickerson F, Jones-Brando L, Ford G, Genovese G, Stallings C, Origoni A, O'Dushlaine C, Katsafanas E, Sweeney K, Khushalani S, Yolken R. Schizophrenia is associated with an aberrant immune response to Epstein-Barr virus. *Schizophr Bull*. 2019;45:1112–9. <https://doi.org/10.1093/schbul/sby164>.
- Dickerson F, Katsafanas E, Origoni A, Squire A, Khushalani S, Newman T, Rowe K, Stallings C, Savage CLG, Sweeney K, Nguyen TT, Breier A, Goff D, Ford G, Jones-Brando L, Yolken R. Exposure to Epstein-Barr virus and cognitive functioning in individuals with schizophrenia. *Schizophr Res*. 2021;228:193–7. <https://doi.org/10.1016/j.schres.2020.12.018>.
- Dickerson F, Schroeder JR, Nimgaonkar V, Gold J, Yolken R. The association between exposure to herpes simplex virus type 1 (HSV-1) and cognitive functioning in schizophrenia: a meta-analysis. *Psychiatry Res*. 2020;291:113157. <https://doi.org/10.1016/j.psychres.2020.113157>.
- Endres D, Huzly D, Dersch R, Stich O, Berger B, Schuchardt F, Perlov E, Venhoff N, Hellwig S, Fiebich BL, Erny D, Hottenrott T, Tebartz van Elst L. Do patients with schizophreniform and bipolar disorders show an intrathecal, polyspecific, antiviral immune response? a pilot study. *Fluids Barriers CNS*. 2017;14:34. <https://doi.org/10.1186/s12987-017-0082-1>.
- Endres D, Meixensberger S, Dersch R, Feige B, Stich O, Venhoff N, Matysik M, Maier SJ, Michel M, Runge K, Nickel K, Urbach H, Domschke K, Prüss H, Tebartz van Elst L. Cerebrospinal fluid, antineuronal autoantibody, EEG, and MRI findings from 992 patients with schizophreniform and affective psychosis. *Transl Psychiatry*. 2020;10:279. <https://doi.org/10.1038/s41398-020-00967-3>.
- Flegr J, Prandota J, Sovičková M, Israili ZH. Toxoplasmosis—a global threat correlation of latent toxoplasmosis with specific disease burden in a set of 88 countries. *PLoS ONE*. 2014. <https://doi.org/10.1371/journal.pone.0090203>.
- Fuglewick AJ, Piotrowski P, Stodolak A. Relationship between toxoplasmosis and schizophrenia: a review. *Adv Clin Exp Med*. 2017. <https://doi.org/10.17219/acem/61435>.
- Gieß RM, Pfuhl C, Behrens JR, Rasche L, Freitag E, Khalighy N, Otto C, Wuerfel J, Brandt AU, Hofmann J, Eberspächer B, Bellmann-Strobl J, Paul F, Ruprecht K. Epstein-Barr virus antibodies in serum and DNA load in saliva are not associated with radiological or clinical disease activity in patients with early multiple sclerosis. *PLoS ONE*. 2017;12:e0175279. <https://doi.org/10.1371/journal.pone.0175279>.
- Gotlieb-Stematsky T, Zonis J, Arlazoroff A, Mizes T, Sigal M, Szekeley AG. Antibodies to Epstein-Barr virus, herpes simplex type 1, cytomegalovirus and measles virus in psychiatric patients. *Adv Virol*. 1981;67:333–9. <https://doi.org/10.1007/BF01314836>.
- Hottenrott T, Dersch R, Berger B, Rauer S, Eckenweiler M, Huzly D, Stich O. The intrathecal, polyspecific antiviral immune response in neurosarcoidosis, acute disseminated encephalomyelitis and autoimmune encephalitis compared to multiple sclerosis in a tertiary hospital cohort. *Fluids Barriers CNS*. 2015;12:27. <https://doi.org/10.1186/s12987-015-0024-8>.
- Hottenrott T, Schorb E, Fritsch K, Dersch R, Berger B, Huzly D, Rauer S, Tebartz van Elst L, Endres D, Stich O. The MRZ reaction and a quantitative intrathecal IgG synthesis may be helpful to differentiate between primary central nervous system lymphoma and multiple sclerosis. *J Neurol*. 2018;265:1106–14. <https://doi.org/10.1007/s00415-018-8779-x>.
- Jones-Brando L. Drugs used in the treatment of schizophrenia and bipolar disorder inhibit the replication of *Toxoplasma gondii*. *Schizophr Res*. 2003;62:237–44. [https://doi.org/10.1016/s0920-9964\(02\)00357-2](https://doi.org/10.1016/s0920-9964(02)00357-2).
- Khandaker GM, Stochl J, Zammit S, Lewis G, Jones PB. Childhood Epstein-Barr Virus infection and subsequent risk of psychotic experiences in adolescence: a population-based prospective serological study. *Schizophr Res*. 2014;158:19–24. <https://doi.org/10.1016/j.schres.2014.05.019>.

24. Khandaker GM, Zimbron J, Lewis G, Jones PB. Prenatal maternal infection, neurodevelopment and adult schizophrenia: a systematic review of population-based studies. *Psychol Med.* 2013;43:239–57. <https://doi.org/10.1017/S0033291712000736>.
25. King DJ, Cooper SJ, Earle JA, Martin SJ, McFerran NV, Wisdom GB. Serum and CSF antibody titres to seven common viruses in schizophrenic patients. *Br J psychiatry.* 1985;147:145–9. <https://doi.org/10.1192/bjp.147.2.145>.
26. Kolcava J, Hulova M, Benesova Y, Bednarik J, Stourac P. The value of anti-JCV antibody index assessment in multiple sclerosis patients treated with natalizumab with respect to demographic, clinical and radiological findings. *Multiple Scler Relat Disord.* 2019;30:187–91. <https://doi.org/10.1016/j.msard.2019.02.019>.
27. Kuzior H, Fiebich BL, Yousif NM, Saliba SW, Ziegler C, Nickel K, Maier SJ, Süß P, Runge K, Matysik M, Dersch R, Berger B, Robinson T, Venhoff N, Kessler F, Blank T, Domschke K, Tebartz van Elst L, Endres D. Increased IL-8 concentrations in the cerebrospinal fluid of patients with unipolar depression. *Compr Psychiatry.* 2020;102: 152196. <https://doi.org/10.1016/j.comppsy.2020.152196>.
28. Leweke FM, Gerth CW, Koethe D, Klosterkötter J, Ruslanova I, Krivogorsky B, Torrey EF, Yolken RH. Antibodies to infectious agents in individuals with recent onset schizophrenia. *Eur Arch Psychiatry Clin Neurosci.* 2004;254:4–8. <https://doi.org/10.1007/s00406-004-0481-6>.
29. Marder SR, Cannon TD. Schizophrenia. *N Engl J Med.* 2019;381:1753–61. <https://doi.org/10.1056/NEJMr1808803>.
30. Martelius T, Lappalainen M, Palomäki M, Anttila V-J. Clinical characteristics of patients with Epstein Barr virus in cerebrospinal fluid. *BMC Infect Dis.* 2011;11:281. <https://doi.org/10.1186/1471-2334-11-281>.
31. Michel M, Fiebich BL, Kuzior H, Meixensberger S, Berger B, Maier S, Nickel K, Runge K, Denzel D, Pankratz B, Schiele MA, Domschke K, Tebartz van Elst L, Endres D. Increased GFAP concentrations in the cerebrospinal fluid of patients with unipolar depression. *Transl Psychiatry.* 2021. <https://doi.org/10.1038/s41398-021-01423-6>.
32. Mohagheghi M, Eftekharian MM, Taheri M, Alikhani MY. Determining the IgM and IgG antibodies titer against HSV1, HSV2 and CMV in the serum of schizophrenic patients. *Hum Antibodies.* 2018;26:87–93. <https://doi.org/10.3233/HAB-170325>.
33. Mortensen PB, Pedersen CB, Hougaard DM, Nørgaard-Petersen B, Mors O, Børglum AD, Yolken RH. A Danish national birth cohort study of maternal HSV-2 antibodies as a risk factor for schizophrenia in their offspring. *Schizophr Res.* 2010;122:257–63. <https://doi.org/10.1016/j.schres.2010.06.010>.
34. de Oliveira MF, Yamashita RHG, Boa Sorte AA, Rotta JM, Norremose KA, Teixeira MJ, Pinto FCG. Psychiatric symptoms are frequent in idiopathic intracranial hypertension patients. *Neurosurg Rev.* 2021;44:1183–9.
35. Pollak TA, Lennox BR, Müller S, Benros ME, Prüss H, Tebartz van Elst L, Klein H, Steiner J, Frodl T, Bogerts B, Tian L, Groc L, Hasan A, Baune BT, Endres D, Haroon E, Yolken R, Benedetti F, Halaris A, Meyer JH, Stassen H, Leboyer M, Fuchs D, Otto M, Brown DA, Vincent A, Najjar S, Bechter K. Autoimmune psychosis: an international consensus on an approach to the diagnosis and management of psychosis of suspected autoimmune origin. *Lancet Psychiat.* 2020;7:93–108. [https://doi.org/10.1016/S2215-0366\(19\)30290-1](https://doi.org/10.1016/S2215-0366(19)30290-1).
36. Prandota J, Gryglas A, Fuglewicz A, Zesławska-Falańczyk A, Ujma-Czapka B, Szenborn L, Mierzwa J. Recurrent headaches may be caused by cerebral toxoplasmosis. *World J Clin Pediatr.* 2014;3:59–68. <https://doi.org/10.5409/wjcp.v3.i3.59>.
37. R Core Team, 2021. R: A Language and environment for statistical computing. vienna, austria. <https://www.R-project.org/>.
38. Reiber H, Peter JB. Cerebrospinal fluid analysis: disease-related data patterns and evaluation programs. *J Neurol Sci.* 2001;184:101–22. [https://doi.org/10.1016/S0022-510X\(00\)00501-3](https://doi.org/10.1016/S0022-510X(00)00501-3).
39. Runge K, Fiebich BL, Kuzior H, Saliba SW, Yousif NM, Meixensberger S, Nickel K, Denzel D, Schiele MA, Maier SJ, Berger B, Dersch R, Domschke K, Tebartz van Elst L, Endres D. An observational study investigating cytokine levels in the cerebrospinal fluid of patients with schizophrenia spectrum disorders. *Schizophr Res.* 2021;231:205–13. <https://doi.org/10.1016/j.schres.2021.03.022>.
40. Runge K, Tebartz van Elst L, Maier S, Nickel K, Denzel D, Matysik M, Kuzior H, Robinson T, Blank T, Dersch R, Domschke K, Endres D. cerebrospinal fluid findings of 36 adult patients with autism spectrum disorder. *Brain Sci.* 2020;10:355. <https://doi.org/10.3390/brainsci10060355>.
41. Rush AJ. Handbook of psychiatric measures. 1st ed. Washington, DC: American Psychiatric Association; 2000.
42. Schizophrenia Working Group of the Psychiatric Genomic Consortium, Ripke S, Neale BM, Corvin A, Walters JTR, Farh KH, Holmans PA, et al. Biological insights from 108 schizophrenia associated genetic loci. *Nature.* 2014. <https://doi.org/10.1038/nature13595>.
43. Shamier MC, Bogers S, Yusuf E, van Splunter M, ten Berge JCEM, Titulaer M, van Kampen JJA, GeurtsvanKessel CH. The role of antibody indexes in clinical virology. *Clinical Microbiol Infect.* 2021. <https://doi.org/10.1016/j.cmi.2021.03.015>.
44. Sherchan R, Shrestha J, Omotosho YB, Dyatlova N, Nepomuceno JS. Herpes Simplex Virus-2 Meningitis Masquerading as Pseudotumor Cerebri. *Cureus.* 2021;13:e15764. <https://doi.org/10.7759/cureus.15764>.
45. Srikanth S, Ravi V, Poornima K, Shetty KT, Gangadhar BN, Janakiramaiah N. Viral antibodies in recent onset, nonorganic psychoses: correspondence with symptomatic severity. *Biol Psychiat.* 1994;36:517–21. [https://doi.org/10.1016/0006-3223\(94\)90615-7](https://doi.org/10.1016/0006-3223(94)90615-7).
46. Stepanova EV, Kondrashin AV, Sergiev VP, Morozova LF, Turbina NA, Maksimova MS, Romanov DV, Kinkulkin MA, Lazareva AV, Morozov EN. Toxoplasmosis and mental disorders in the Russian Federation (with special reference to schizophrenia). *PLoS ONE.* 2019;14: e0219454. <https://doi.org/10.1371/journal.pone.0219454>.
47. Stich O, Andres TA, Gross CM, Gerber SI, Rauer S, Langosch JM. An observational study of inflammation in the central nervous system in patients with bipolar disorder. *Bipolar Disord.* 2015;17:291–302. <https://doi.org/10.1111/bdi.12244>.
48. Tanaka T, Matsuda T, Hayes LN, Yang S, Rodriguez K, Severance EG, Yolken RH, Sawa A, Eaton WW. Infection and inflammation in schizophrenia and bipolar disorder. *Neurosci Res.* 2017;115:59–63. <https://doi.org/10.1016/j.neures.2016.11.002>.
49. Tebartz van Elst L. Vom Anfang und Ende der Schizophrenie: Eine neuropsychiatrische Perspektive auf das Schizophrenie-Konzept. Stuttgart: Kohlhammer Verlag; 2017. p. 1255.
50. Tedla Y, Shibre T, Ali O, Tadele G, Woldeamanuel Y, Asrat D, Aseffa A, Mihret W, Abebe M, Alem A, Medhin G, Habte A. Serum antibodies to toxoplasma gondii and Herpesviridae family viruses in individuals with schizophrenia and bipolar disorder: a case-control study. *Ethiop Med J.* 2011;49:211–20.
51. Torrey EF, Bartko JJ, Yolken RH. Toxoplasma gondii and other risk factors for schizophrenia: an update. *Schizophr Bull.* 2012;38:642–7. <https://doi.org/10.1093/schbul/sbs043>.
52. Torrey EF, Leweke MF, Schwarz MJ, Mueller N, Bachmann S, Schroeder J, Dickerson F, Yolken RH. Cytomegalovirus and schizophrenia. *CNS Drugs.* 2006;20:879–85. <https://doi.org/10.2165/00023210-200620110-00001>.
53. Torrey EF, Peterson MR, Brannon WL, Carpenter WT, Post RM, van Kammen DP. Immunoglobulins and viral antibodies in psychiatric patients. *Br J psychiatry.* 1978;132:342–8. <https://doi.org/10.1192/bjp.132.4.342>.
54. Torrey EF, Yolken RH, Winfrey CJ. Cytomegalovirus antibody in cerebrospinal fluid of schizophrenic patients detected by enzyme immunoassay. *Science.* 1982. <https://doi.org/10.1126/science.6281883>.
55. Tucker JD, Bertke AS. Assessment of cognitive impairment in HSV-1 positive schizophrenia and bipolar patients: systematic review and meta-analysis. *Schizophr Res.* 2019;209:40–7. <https://doi.org/10.1016/j.schres.2019.01.001>.
56. Tzartos JS, Khan G, Vossenkamper A, Cruz-Sadaba M, Lonardi S, Sefia E, Meager A, Elia A, Middeldorp JM, Clemens M, Farrell PJ, Giovannoni G, Meier U-C. Association of innate immune activation with latent Epstein-Barr virus in active MS lesions. *Neurology.* 2012;78:15–23. <https://doi.org/10.1212/WNL.0b013e31823ed057>.
57. Wickham H. ggplot2: Elegant graphics for data analysis. Cham: Springer; 2016. p. 260.
58. Wilking H, Thamm M, Stark K, Aebischer T, Seeber F. Prevalence, incidence estimations, and risk factors of Toxoplasma gondii infection in Germany: a representative, cross-sectional, serological study. *Sci Rep.* 2016;6:22551. <https://doi.org/10.1038/srep22551>.
59. de Witte LD, van Mierlo HC, Litjens M, Klein HC, Bahn S, Osterhaus AD. The association between antibodies to neurotropic pathogens and schizophrenia: a case-control study. *NPJ Schizophr.* 2015;1:15041. <https://doi.org/10.1038/npjischz.2015.41>.
60. Xiao J, Prandovszky E, Kannan G, Pletnikov MV, Dickerson F, Severance EG, Yolken RH. Toxoplasma gondii: biological parameters of the connection

to schizophrenia. *Schizophr Bull.* 2018;44:983–92. <https://doi.org/10.1093/schbul/sby082>.

61. Yolken R, Torrey EF, Dickerson F. Evidence of increased exposure to *Toxoplasma gondii* in individuals with recent onset psychosis but not with established schizophrenia. *PLoS Negl Trop Dis.* 2017;11: e0006040. <https://doi.org/10.1371/journal.pntd.0006040>.
62. Yolken RH, Torrey EF. Viruses, schizophrenia, and bipolar disorder. *Clin Microbiol Rev.* 1995;8:131–45. <https://doi.org/10.1128/CMR.8.1.131>.
63. Yolken RH, Torrey EF. Are some cases of psychosis caused by microbial agents? a review of the evidence. *Mol Psychiat.* 2008;13:470–9. <https://doi.org/10.1038/mp.2008.5>.

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